CH_2CH_2OCO), 4.5 (m, 4 H, CH_2OCO), 7.6 (t, 2 H, ArH), 8.2 (d. 2 H, ArH), 8.3 (d, 2 H, ArH); mass spectrum, m/e M⁺ 500. Anal. Calcd for $C_{24}H_{20}O_8S_2$: C, 57.59; H, 4.03; S, 12.81. Found: C, 57.53; H, 3.91; S, 12.78.

7,7'-Bis(((2-((2-(methoxyethylene)oxy)ethylene)oxy)ethylene)oxy)carbony)thioindigo (T-2). The synthetic procedure was the same as for T-1. Triethylene glycol monomethyl ether was used instead of ethylene glycol monomethyl ether: ¹H NMR (Me₂SO-d₆) 3.3 (s, 6 H, CH₃), 3.5 (m, 16 H, CH₂), 3.8 (m, 4 H, CH₂CH₂OCO), 4.5 (m, 4 H, CH₂OCO), 7.6 (t, 2 H, ArH), 8.2 (d, 2 H, ArH), 8.3 (d, 2 H, ArH); mass spectrum, m/e M⁺ 676. Anal. Calcd for C₃₂H₃₆O₁₂S₂: C, 56.79; H, 5.36; S, 9.48. Found: C, 56.76; H, 5.53; S, 9.55.

Photoisomerization. A xenon lamp (Osram 450 W) was used as a light source. Monochromatic light of 550 and 450 nm was obtained by passing the light through a band-pass interference filter (Toshiba KL-55 or KL-45) and a cutoff filter (Toshib VY-42). Lines of the Ar ion laser (Spectra-Physics 165) of 488 and 529 nm were also used to isomerize T-1 from the cis to the trans and from the trans to the cis form, respectively, in a single test tube or in a H-type ion transport cell. A laser beam is convenient for irradiation to the limited area. When used for the ion transport experiments, the laser beam irradiated a small area just below the aqueous-organic phase boundary.

Solvent Extraction. Equal volumes (15 mL) of benzene (or 1,2-dichloroethane) containing $4 \times 10^{-5} \text{ mol}/1 \text{ T-1}$ or T-2 and an aqueous solution containing $1 \times 10^{-2} \text{ mol}/1$ metal nitrate and $2 \times 10^{-5} \text{ mol}/1$ picric acid was vigorously agitated for 30 min with a shaker (Yamato SA-31). T-1 and T-2 were irradiated with 450- or 550-nm light for 10 min prior to the extraction experiments. A similar extraction was performed with pure benzene (or 1,2-dichloroethane). The extractability was determined from the difference between these two absorbances of metal picrate in the aqueous phase. All the extractions were conducted at 25 °C.

Ion Transport across a Liquid Membrane. A H-type cell containing 40 mL of organic liquid membrane (1,2-dichloroethane) and two aqueous phases (15 mL each) was used. The cell was immerced in a water bath thermostated at 25 °C. The liquid membrane was stirred gently. Dissolved in the liquid membrane was 4×10^{-5} mol/1 T-1. Aqueous phase I contained 1×10^{-2} mol/1 AgNO₃ and 2×10^{-5} mol/1 picric acid, and phase II was pure water. The ion transport was followed by measuring the decrease of the absorption of metal picrate in phase I and the increase in phase II.

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On a Tandem 1,2-Elimination/[1,7]-Sigmatropic Shift: Synthesis of Double Bond Shifted Isomers of Vitamin A

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Abstract: The syntheses of four geometrical isomers of 18,14-retro-retinol (7a-10a) and two dehydro derivatives (31b and 32b) are described. Treatment of the *cis*-benzoate 19 with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) afforded an 80% yield of the trienyne 20a. Evidence for the formation of the latter by an initial 1,2-elimination to afford the putative 21 followed by a [1,7]-sigmatropic shift is provided. Thus, treatment of the pentadeuteriobenzoate 23 under identical conditions afforded 25, in which a single deuterium atom is incorporated at the C_3 , methyl position by way of an intramolecular process. Palladium-catalyzed coupling of the Grignard salt of 20a (20f) with (*E*)-bromide 30a followed by deprotection afforded dehydro-retro-retinol 31b. The latter was catalytically reduced to the (10Z)-retro-retinol 8a. A similar sequence applied to the (2)-bromide 30b afforded 32b and the (10Z,12Z)-retro-retinol 10a. A standard three-step sequence was used to convert 20a to the tetraenal 33b, which was condensed in a Wittig reaction with ylide 35 to afford after separation the *all-trans*- and (12Z)-retro-retinols, 7a and 9a, respectively.

A significant correlation between the role of retinoids (vitamin A) in controlling differentiation of epithelial cells and the development of malignancy in epithelial tissues has been clearly established.¹ The natural vitamin A (1) exhibits therapeutic and prophylactic activity in certain types of carcinomas but unfortunately causes toxic liver damage due to excessive localization in the liver. In contrast, the unnatural geometric isomer 13-cis-



retinoic acid (2) is far less toxic yet possesses similar biological efficacy in controlling cell differentiation.^{1,2} Success of this sort has spurred a search for increasingly more effective retinoidal

anticancer drugs.³ The signal importance of retinoids in photobiological research (energy transduction and vision)⁴ and acne therapy⁵ provides further impetus for synthetic endeavors in the vitamin A field.

In the case of compounds which show toxic side effects, acceptable therapeutic indexes can still be obtained by designing analogues which show greatly enhanced beneficial activity. Although exactly how a retinoid such as 2 exerts its various biological effects is unclear, it would be quite plausible that it does so by binding to one or more proteins in one topologically specific form or another. For example, 13-cis-retinoic acid (2) may bind to a putative receptor in a side chain conformation, such as 3-6 or one

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of a number of other rotamers, which is not necessarily the lowest energy conformation. Thus, a conformationally locked analogue



which mimics the "active topology" would be expected to bind preferentially and exhibit enhanced biological activity. The systematic fusion of rings to the retinoid side chain has produced conformationally locked analogues which in some cases have been extraordinarily active.⁶ We envisage an alternative mode of conformational locking where the interchange of double and single bond positions along the side chain would produce the locked equivalents of various conformations of different geometric isomers. With this concept in mind, we became interested in synthesizing and studying the polyenes 7-10 which in their most extended conformations would be topological mimics of the conformational isomers of 13-cis-retinoic acid 3-6, respectively.



g, x=CH2OH; b, X=COOH

The 18,14-retro-retinois 7-10 possess the partial chromophore of the hexaenamine 11, a substance hypothesized as an intermediate in the visual process.⁷ Thus, these retro-retinoids are also of interest as possible synthetic precursors of 11 and its geometric isomers.

In previous synthetic endeavors in the retinoid area, we utilized thermally induced [1,5]-sigmatropic hydrogen shifts in 9,10-allenes of the type 12 to obtain geometrically isomeric 11-cis-retinoids 13 and various conformationally locked analogues.⁸ While a similar [1,5]-hydrogen migration of the corresponding 7-cis-9,10-allene 14 would result in various 7-cis,11-cis-retinoids 15, we anticipated instead that 14 would preferentially undergo a



thermal [1,7]-sigmatropic hydrogen shift from C_{18} to C_{10} to give 16 and hence provide us initial access to the retro-retinoid series. However, in studies directed toward this end, we observed that the model compounds 17 did not undergo the desired [1,7]-hydrogen shift but instead underwent very facile six-electron electrocyclizations to give the bicyclic products 18,9 indicating that the allene approach to retro-retinoids was probably not a viable one. The unexpected discovery was made, however, that the cis-benzoate 19, utilized previously9 in preparing some derivatives of 18 via 17, could be transformed into trienyne 20a (vide infra), which possessed the desired 18-retro skeleton. It is the purpose



of this article to describe the synthesis of the double bond shifted isomers 7a-10a and related substances from the trienyne 20a and to show that the latter is in fact produced by a most unusual tandem 1,2-elimination/[1,7]-sigmatropic shift via 21 from 19.

Results and Discussion

The base-induced (DBU, refluxing acetonitrite) elimination of benzoic acid from cis-benzoate 199 surprisingly afforded a single

⁽⁶⁾ For leading references, see: Dawson, M. I.; Hobbs, P. D.; Chan, R.

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hydrocarbon (80%) as ascertained by ¹H and ¹³C NMR spectroscopy as well as chromatography. Its ¹H NMR clearly revealed the presence of a terminal vinyl group consistent with either 20a or 21, but a more detailed analysis revealed it to be the former. The appearance of appropriate allylic couplings (between $H_{2'}$ and the C_{3'} CH₃, $J \sim 1.4$ Hz; H₃ and the Z hydrogen of the exocyclic methylene group, J ~ 1.4 Hz)¹⁰ and the vicinal s-trans coupling (between $H_{1'}$ and $H_{2'}$ of $J \sim 10.8 \text{ Hz})^{11}$ provide the most convincing evidence for structure 20a rather than 21. The formation of only 20a, to the exclusion of its three other possible geometric isomers and of structural isomers of the type 22, cannot satisfactorily be rationalized on the basis of a direct 1,6-elimination of benzoic acid. However, a 1,2-elimination of benzoic acid leading to the initial formation of 21 followed by a subsequent [1,7]sigmatropic hydrogen shift would readily explain the formation of a single, more stable, linearly conjugated trienyne (20a) from the cross-conjugated (21) system.

Since the geometry assigned to the trienyne 20a (and also to the retro-retinots and intermediates subsequently derived from 20a) is based on the above mechanistic hypothesis, it was imperative that we unequivocally establish this mechanistic scheme. All attempts made to isolate the putative intermediate 21 by inducing the reaction of 19 at lowered temperatures were unsuccessful. We envisaged that reaction of the deuteriobenzoate 23 under elimination conditions would, if the postulated mechanism were operative, lead through the intermediacy of 24 to 25 with the incorporation of a single deuterium atom at the $C_{3'}$ CH₃ and net retention of deuterium content. Alternatively, if a direct



1,6-elimination were to occur, reaction of 23 would likely result in 26 with a net loss of a single deuterium. The desired benzoate 23 was synthesized starting from the known pentadeuterio- β cyclocitral 27,12 which upon condensation with diethyl (cyanomethyl)phosphonate13 followed by treatment of the resultant diene nitrile 28 with methyllithium gave upon acidic workup¹⁴ the pentadeuterio- β -ionone 29. The latter (29) was then converted to 23 by a sequence identical with that used for the synthesis of 19.9 Treatment of 23 under conditions identical with those used for the synthesis of 20a afforded a single hydrocarbon product 25 in which we could demonstrate clearly by ¹H and ¹³C NMR the presence of a single deuterium in the $C_{3'}$ methyl group. Mass spectroscopic analysis of 23 and 25 also revealed that the conversion proceeds with retention of deuterium content. These data



clearly indicate that the formation of 25 (and 20a) must proceed by a [1,7]-sigmatropic hydrogen shift of the cross-conjugated 24 (or 21 in the undeuterated case) and hence confirms the stereochemistry assigned to 25 (and 20a).

Parenthetically, it is interesting to note at this stage the dichotomy in reaction pathway of the putative intermediates 17 and 21. While the 7-cis-vinylallenes 17 react exclusively by six-electron electrocyclization, the corresponding triene 21 reacts by a [1,7]-hydrogen shift. Apparently, the extraordinarity facile sixelectron electrocyclization pathway accessible to vinylallenes of the type 17 results from the central carbon of the allene being a site of minimal steric demand. The introduction of even a small amount of steric hindrance in the form of the hydrogens in the terminal vinyl group of the putative 21 results in a total crossover to the [1,7]-hydrogen shift pathway. However, at higher temperatures, electrocyclizations will very likely occur in the latter case as exemplified by the thermal behavior of β -carotene.¹³

It was envisaged that cross-coupling¹⁵ of the zinc salt 20b derived from the trienyne 20a with the protected vinyl bromides **30a** (E) and **30b** (Z) in the presence of pathadium catalyst would lead to tetraenynes 31 and 32, respectively. Subsequent catalytic



reduction should afford two (8a and 10a, respectively) of the four retro-retinols. This process, reported to be applicable to alkenyl bromides as well as iodides,^{15a} was not successful. The zinc salt 20b also failed to react when 30 was replaced by iodobenzene in order to produce the phenyl derivative 20c.^{15b} However the terminal copper derivative 20d derived from 20a reacted with iodobenzene in the presence of catalyst to afford an 83% yield of 20c but failed to react with 30a, 30b, or bromobenzene.¹⁶ The iodoacetylene 20e also failed to reat with the copper salt derived from the bromide 30a.¹⁷ The successful route¹⁸ entailed conversion of 20a to its magnesium salt 20f, which in the presence of catalytic amounts of tetrakis(triphenylphosphine)palladium reacted with 30a (benzene; 2 h, room temperature, and 10 h, 60 °C) to give after deprotection [$(n-Bu)_4$ NF, THF] pure E isomer **31b** in 69% yield. Reaction of the (Z)-bromide 30b in a similar manner afforded the Z isomer 32b contaminated by E isomer 31b, which after separation were obtained in 63% and 6.2% yields, respectively.

⁽¹⁰⁾ The triene portion of the molecule exhibits a ¹H NMR pattern very similar to that of vitamin D₃ and the even more closely related 4,4-di-methyl-vitamin D₃. See: (a) Wing, R. M.; Okamura, W. H.; Rego, A.; Pirio, M. R.; Norman, A. W. J. Am. Chem. Soc. **1975**, *97*, 4980. (b) Berman, E.; Friedman, N.; Mazur, Y.; Sheves, M. Ibid. 1978, 100, 5626.

⁽¹¹⁾ The observed cis vicinal coupling in 19, a model for the putative 21, is $J \sim 13.5$ Hz. The value of 10.8 Hz observed for the hydrocarbon assigned 20a would appear to also be at variance with that expected for 21.

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Lindlar reduction of **31b** and **32b** afforded after high-pressure LC purification the desired Δ^{10} -cis-retro-retinols **8a** (58%) and **10a** (61%), respectively.

The Δ^{10} -trans isomers 7a and 9a were prepared form 20a by a chain extension procedure. Reaction of the lithium salt of 20a with paraformaldehyde¹⁹ afforded the propargyl alcohol 20g (82%), which was reduced (LiA1H₄, THF)²⁰ to the allylic alcohol 33a (~quantitative) and then the latter oxidized (MnO₂, petroleum ether) to aldehyde 33b (81%). Wittig condensation of 33b with ylide 35 (Scheme I),²¹ prepared in situ from 34,²² afforded after separation the desired Δ^{10} -trans isomers 9a (20%) and 7a (30%).

In summary, we have developed a simple route to 18,14retro-retinols through the intermediacy of the novel trienyne 20a. The unusual formation of 20a was clearly established as a tandem β -elimination/[1,7]-sigmatropic shift by appropriate labeling studies. The new 18,14-retro-retinols reported here (7a-10a), besides being promising candidates for evaluation as potential chemotherapeutic and chemoprophylactic agents, provide prototypes for preparing related end group modified analogues such as the acids 7b-10b as well as 11 for future biological evaluation.

Experimental Section

General, ¹H nuclear magnetic resonance spectra (¹H NMR), 90-MHz Varian EM-390 or 200-MHz Jeol FX-200 spectrometers, were performed with deuteriochloroform (CDCl₃) with tetramethylsilane (TMS) as the internal standard; chemical shifts are given in delta (δ) values, and the coupling constants (J) are given in hertz (Hz). ¹³C nuclear magnetic resonance spectra (¹³C NMR), 50.4-MHz Jeol FX-200 spectrometer were performed with deuteriochloroform (CDCl₃) as the solvent and internal standard; chemical shifts are given in delta (δ) values and coupling constants (J) in hertz (Hz). Infrared spectra (IR), Perkin-Elmer Model 283 grating spectrophotometer, 0.1 mm NaCl plates, were performed in CDCl₃ or CCl₄ solutions as indicated. Mass spectra low resolution (MS) and mass spectra exact mass (MS exact mass), m/e (rel intensity), 70 eV, were performed on Varian MAT 711 and Kratos MS-50 and MS-80 mass spectrometers (the National Science Foundation Midwest Center for Mass Spectrometry). Ultraviolet spectra (UV) were performed on a Cary 17 or Cary 219D (Varian) spectrophotometer with 95% ethanol as the solvent unless otherwise indicated.

Air sensitive materials were generally stored under nitrogen in a -80 °C freezer, and reactions involving organometallic materials were performed under an atmosphere of dry nitrogen or argon. Dry ether or THF (tetrahydrofuran) refers to reagent grade material freshly distilled from LiAlH₄ under nitrogen. Skellysolve B, hexanes, and lbpe (low-boiling petroleum ether, bp 30-60 °C) were distilled from CaH₂ prior to use. Pyridine was distilled from CaH₂ or KOH and stored over 4-Å molecular sieves. Kugelrohr distillation boiling points (bp) refer to the external air bath temperature.

High-pressure liquid chromatography (high-pressure LC) was performed using a Waters 6000A solvent delivery system equipped with a U6K injector and dual-detector system (M450 variable wavelength UV and R401 refractive index detectors). A Whatman M9 10/50 Partisil (10- μ m particle size, 9.4 mm i.d. \times 50 cm) column was used except as noted. All chromatographic solvents were distilled prior to use. Solvents and solvent mixtures were vacuum-filtered through a 0.45- μ m Millipore filter and vacuum degassed immediately prior to use. Ordinary column chromatography was performed using J. T. Baker silica gel (60-200 mesh). Thin-layer chromatography (TLC) was performed using precoated plates (silica gel 60 F-254 from MCB-Merck).

(6Z, 8E, 10E, 12E)- and (6Z, 8E, 10E, 12Z)-18, 14-retro-Retinol (7a and 9a, respectively). Normal butyllithium (0.52 mL, 1.6 M, 0.83 mmol) was added dropwise to a stirred suspension of phosphonium salt 34 (121 mg, 0.3 mmol) in dry THF (2.5 mL) under argon at room temperature. The initial yellow color turned a dark brick red color after 10 min. After 15 min, iodomethane (86 mg, 0.6 mmol) was added dropwise, producing an immediate white precipitate. After stirring for 15 min, *n*-BuLi (0.27 mL, 1.6 M, 0.43 mmol) was added, producing again a deep brick red coloration. After 15 min, the aldehyde 33b (58 mg, 0.25 mmol) in dry THF (1 mL) was introduced and the mixture was stirred for 1 h. Water was added, and the mixture was extracted with ether (3 × 15 mL). The dried ether layer was evaporated to dryness and then the residue was filtered with ether through a short column of silica gel. Removal of ether left a residue which was subjected to purification by high-pressure LC (Whatman M9 10/50 partisil column, 25% ethyl acetate/Skellysolve B, 4 mL/min flow rate). The pure products were eluted as follows: the less polar 12Z isomer A, 9a (14.6 mg, 20.4%); an unidentified substance (7.2 mg) with a UV λ_{max} 289 nm (similar to tetraenol 20a); and finally the more polar 12E isomer B, 7a (21.2 mg, 29.6%).

Retinol 7a: ¹H NMR (200 MHz) δ 1.10 (C_{16,17} 2CH₃, s), 1.82 (C₂₀ CH₃, d, $J \sim 1.2$ Hz), 1.92 (C₁₉ CH₃, d, $J \sim 1.2$ Hz), 2.22 (C₄ CH₂, m), 2.36 (C₁₄ CH₂, t, $J \sim 6.0$ Hz), 3.71 (C₁₅ CH₂, t, $J \sim 6.0$ Hz), 4.69 (C₁₈ CH₂, d, $J \sim 3$ Hz), 5.10 (C₁₈ CH₂, dt, $J \sim 3$, 1.2 Hz), 5.99 (H₁₂, br d, $J \sim 10.3$ Hz), 6.25 (H₇, d, $J \sim 11.4$ Hz), 6.26 (H₁₀, d, $J \sim 14.5$ Hz), 6.41 (H₁₁, dd, $J \sim 14.5$, 10.3 Hz), 6.49 (C_{16,17}; UV (95% EtOH) λ_{max} 333 nm (ϵ 40 500), 318 (ϵ 46 000), 306 (sh, ϵ 34 800); IR (CCl₄) ν_{max} 3360 (s, OH), 2950, 2880, 1550, 1245, 1040, 855, 760 cm⁻¹; MS exact mass, *m/e* 286 (.291 (calcd for C₂₀H₃₀O, 286.2298); MS, *m/e* 287 (1, M + 1), 286 (7, M), 181 (34), 159 (base), 145 (28), 143 (35), 135 (26), 133 (30), 131 (73), 129 (30), 128 (26), 119 (67), 115 (29), 105 (54), 95 (35), 93 (37), 91 (51), 81 (29), 79 (29).

Retinol 9a: ¹H NMR (200 MHz) δ 1.10 (C_{16,17} 2CH₃, s), 1.85 (C₂₀ CH₃, s), 1.93 (C₁₉ CH₃, s), 2.23 (C₄ CH₂, m), 2.51 (C₁₄ CH₂, t, $J \sim 6$ Hz), 3.73 (C₁₅ CH₂, br q, $J \sim 6$ Hz), 4.68 (C₁₈ CH_E, d, $J \sim 3$ Hz), 5.09 (C₁₈ CH_Z, dt, $J \sim 3$, 1.5 Hz), 6.09 (H₁₂, br d, $J \sim 10.7$ Hz), 6.24 (H₁₀, d, $J \sim 15.0$ Hz), 6.25 (H₇, d, $J \sim 11.5$ Hz), 6.44 (H₁₁, dd, $J \sim 15.0$, 10.7 Hz), 6.48 (H₈, br d, $J \sim 11.5$ Hz); UV (95% EtOH) λ_{max} 333 nm (ϵ 36 000), 318 (ϵ 40 700), 306 (sh, ϵ 31 500); IR (CCl₄) ν_{max} 3380 (OH), 2960, 2880, 1550, 1245, 1040, 860, 760 cm⁻¹; MS exact mass, m/e 286.2303 (calcd for C₂₀H₃₀O, 286.2298); MS, m/e 287 (5, M + 1), 286 (26, M) 213 (17), 174 (16), 159 (base), 157 (19), 145 (18), 143 (24), 131 (22), 129 (20), 128 (16), 119 (19), 105 (27), 95 (19), 91 (26), 79 (18).

(6Z,8E,10Z,12E)-18,14-retro-Retinol (8a). A low-pressure hydrogenation vessel was charged with Lindlar catalyst (100 mg) in 6 mL of dry benzene and then the apparatus was evacuated and hydrogen admitted to a pressure of ~ 1 atm. The (12E)-acetylenic retinol 31b (14 mg, 0.05 mmol) in 1.5 mL of dry benzene was injected via syringe and then the mixture was stirred at ~ 1 atm (room temperature) for ~ 0.5 h. Hydrogenation ($\sim 2 \text{ mL}$) slows down but does not stop. The vessel was removed, and the mixture was filtered through a celite-sodium sulfate pad and rinsed with ether (10 mL). After removing solvent under vacuum, the residue was passed through a short silica gel column (ether/lbpe, 1:1). Removal of solvent left a residue which was subjected to high-pressure LC purification (Whatman M9 10/50 partisil column, 20% ethyl acetate/Skellysolve B, 4.0 mL/flow rate) to give retro-retinol 8a $\begin{array}{l} (8.2 \text{ mg}, 58\%): \ ^{1}\text{H NMR} \ (200 \text{ MHz}) \ \delta \ 1.10 \ (C_{16,17} \ 2\text{CH}_{3}, \ 6 \ \text{H}, \ \text{s}), \ 1.80 \\ (C_{20} \ \text{CH}_{3}, \ 3 \ \text{H}, \ \text{s}), \ 2.0 \ (C_{19} \ \text{CH}_{3}, \ 3 \ \text{H}, \ \text{s}), \ 2.21 \ (C_{4} \ \text{CH}_{2}, \ 2 \ \text{H}, \ \text{m}), \ 2.37 \end{array}$ $(C_{14} CH_2, 2 H, t, J \sim 6.0 Hz), 3.72 (C_{15} CH_2, t, J \sim 6.0 Hz), 4.69 (C_{18})$ CH_E , 1 H, d, $J \sim 2.3$ Hz), 5.08 (C_{18} CH_Z , 1 H, br m), 5.86 (H_{12} , 1 H, d, $J \sim 11.9$ Hz), 6.10 (H₁₁, 1 H, dd, $J \sim 11.9$ Hz, $J \sim 11.9$ Hz), 6.19 $(H_7, 1 H, d, J \sim 11.2 Hz), 6.42 (H_{10}, 1 H, d, J \sim 11.9 Hz), 6.50 (H_8, J)$ 1 H, d, $J \sim 11.2$ Hz); ¹³C NMR δ 152.6, 146.2, 135.3, 133.3, 129.5, 125.0, 123.2, 116.3, 113.1 (nine peaks, ten olefinic carbons), 60.6, 43.5, 41.5, 38.4, 37.2, 23.3, 17.2, 16.2 ($C_{1-4,14,15,19,20}$), and 27.6 ($C_{16,17}$); UV (95% EtOH) λ_{max} 333 nm (sh, ϵ 29000), 319 (ϵ 34700), 306 (sh, ϵ 29000), 235 (ϵ 12800); IR (CCl₎ ν_{max} 3367, 2930, 1545, 1245, 860 cm⁻¹. MS exact mass, *m/e* 286.2299 (calcd for C₂₀H₃₀O, 286.2298); MS, *m/e* 287 (10, M + 1), 286 (44, M), 271 (44), 213 (44), 185 (41), 175 (26), 173 (40), 171 (29), 159 (94), 157 (48), 145 (46), 143 (55), 133 (43), 131 (48), 129 (35), 121 (31), 119 (56), 117 (30), 109 (29), 107 (38), 105 (86), 95 (base), 93 (55), 91 (79), 83 (31), 81 (89), 79 (53).

(6Z,8E,10Z,12Z)-18,14-retro-Retinol (10a). The (12Z)-acetylenic retinol 32b (12 mg, 0.042 mmol) was hydrogenated with Lindlar catalyst (50 mg) in benzene (4.5 mL total) in precisely the same manner as described for the *E* isomer 31b (see preceding section). The high-pressure LC pure alcohol 10a was obtained in 61% yield (7.2 mg): ¹H NMR (200 MHz) δ 1.10 (C_{16,17} 2CH₃, 6 H, s), 1.88 (C₂₀ CH₃, 3 H, s), 2.01 (C₁₉ CH₃, 3 H, s), 2.20 (C₄ CH₂, 2 H, m), 2.49 (C₁₄ CH₂, 2 H, t, *J* ~ 6.2 Hz), 3.75 (C₁₅ CH₂, 2 H, t, *J* ~ 6.2 Hz), 4.69 (C₁₈ CH_{*E*}, 1 H, d, *J* ~ 1.4 Hz), 5.08 (C₁₈ CH_{*Z*}, 1 H, br m), 5.83 (H₁₂, 1 H, d, *J* ~ 11.6 Hz), 6.48 (H₁₀, 1 H, d, *J* ~ 11.8 Hz), 6.55 (H₈, 1 H, d, *J* ~ 11.6 Hz), 6.48 (H₁₀, 1 H, d, *J* ~ 11.8 Hz), 6.55 (H₈, 1 3.7, 3.6.1, 34.4, 23.2, 22.1, 15.8 (C₁₋₄:14,15,19,20), and 27.2 (C_{16,17}); UV (95% EtOH) λ_{max} 331 nm (sh, ε 32 300), 238 (ε 80, 1550, 1255, 860 cm⁻¹. MS exact mass,

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⁽²⁰⁾ Bates, E. B.; Jones, E. R. H.; Whiting, M. C. J. Chem. Soc. 1954, 1854.

⁽²¹⁾ Bertele, E.; Schudel, P. Helv. Chim. Acta 1967, 50, 2445. (22) Ebata, T.; Mori, K. Agric. Biol. Chem. 1979, 43, 1567.

m/e 286.2298 (calcd for C₂₀H₂₀O, 286.2298); MS, m/e 287 (2, M + 1), 286 (14, M), 201 (86), 175 (41), 173 (32), 159 (86), 157 (32), 147 (32), 145 (44), 143 (41), 133 (56), 131 (46), 121 (40), 119 (56), 109 (51), 107 (44), 105 (90), 97 (60), 95 (95), 93 (52), 91 (79), 85 (40), 83 (70), 81 (base), 79 (52).

1-[(1,1'-Z,2'-E)-3'-Methylpent-2'-en-4'-yn-1'-ylidene]-2-methylene-6,6-dimethylcyclohexane (20a). A mixture of benzoate 19 (1.28 g, 3.98 mmol) and 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU, 0.62 g, 4.0 mmol) in acetonitrile (25 mL) was refluxed for 4 h. The reaction was followed by TLC (8:2 lbpe/ether). The mixture was poured into water (25 mL) and lbpe (25 mL), the organic layer was removed, and the aqueous layer was further extracted with lbpe (25 mL). The combined lbpe extracts were washed successively with saturated aqueous solutions of NH4Cl and NaHCO₃ and then water. After the solution was dried (Na_2SO_4) , the lbpe was removed, and the residue was then chromatographed (silica gel). Elution with lbpe gave the trienyne 20a in 80.5% yield (1.03 g as a viscous oil): ¹H NMR δ 1.03 (C₆ 2CH₃, s), 1.88 (C₃· CH₃, d, $J \sim 1.4$ Hz), 2.83 (1 H, H₅, s), 4.62 (C₂ CH_z, d, $J \sim 2.7$ Hz), 5.02 (C₂ CH_z, d of t, $J \sim 2.7$, 1.4 Hz), 6.05 (1 H, H₁, d, $J \sim 10.8$ Hz), 6.78 (1 H, H₂, d with fine structure, $J \sim 10.8$ Hz); ¹³C NMR δ 154.74 (s), 145.54 (s), a with the structure, $j \sim 10.8 \text{ Hz}$), $\sim 104 \text{ Ke}$ (34, 14, (3), 145.34 (3), 135.28 (d, ${}^{1}\text{J}_{C-H} \sim 156 \text{ Hz}$), 115.48 (d, ${}^{1}\text{J}_{C-H} \sim 157 \text{ Hz}$), 88.23 (C₄', s), 75.63 (C₅', d, ${}^{1}\text{J}_{C-H} \sim 249 \text{ Hz}$), 41.53 (t, ${}^{1}\text{J}_{C-H} \sim 128 \text{ Hz}$), 38.51 (C₆, s), 37.10 (t, ${}^{1}\text{J}_{C-H} \sim 128 \text{ Hz}$), 27.52 (C₆ 2CH₃, q, ${}^{1}\text{J}_{C-H} \sim 125 \text{ Hz}$), 23.19 (t, ${}^{1}\text{J}_{C-H} \sim 128 \text{ Hz}$), 17.26 (C₃' CH₃, q, ${}^{1}\text{J}_{C-H} \sim 129 \text{ Hz}$); IR (CCl₄) ν 3320 (s, C=C-H), 2080 (w, C=C), 903 (s, CH₂), other prominent bands, 2935, 2870, 2850, 1450, 1385, 1205, 635 cm⁻¹; UV (95% EtOH) λ_{max} 280 nm (ϵ 21 200); MS exact mass, m/e 200.1556 (calcd for C₁₅H₂₀, 200.1566); MS, m/e 200 (41, M), 185 (base), 157 (20), 143 (30), 129 (52), 119 (30), 115 (54), 105 (33), 91 (65), 77 (33).

1-[(1,1'-Z,2'-E)-3'-Methyl-5'-phenylpent-2'-en-4'-yn-1'-ylidene]-2methylene-6,6-dimethylcyclohexane (20c). To the trienyne 20a (50 mg, 0.25 mmol) dissolved in dry THF (0.8 mL) was added n-butyllithium (0.2 mL, 1.6 M in hexanes, 0.32 mmol) slowly at 0 °C. After the solution was stirred for 15 min, a suspension of cuprous iodide (42.8 mg, 0.25 mmol) in dry THF (1 mL) was added, and the mixtures were stirred for 20 min at room temperature. The above solution was then added dropwise via cannula to a mixture of tetrakis(triphenylphosphine)palladium (15 mg, 0.0125 mmol) and iodobenzene (51 mg, 0.25 mmol) in dry THF (1 mL) at 0 °C. The mixture was stirred for 2 h at room temperature and then quenched with cold water. Lbpe (20 mL) was added, and the organic layer was separated and passed over a short alumina column. The concentrated residue was chromatographed through silica gel (elution with lbpe) to yield the product 20c (57 mg, 83%): ¹H NMR δ 1.10 (C₆ 2CH₃, s), 1.95 (C₃, CH₃, d, $J \sim 1.5$ Hz), 4.67 (C₂ CH_E, d, $J \sim 2.8$ Hz), 5.08 (C₂ = CH_z, d of t, $J \sim 2.8$, 1.5 Hz), 6.15 (H₁, d, $J \sim 11.2$ Hz), 6.86 (H₂, d of q, $J \sim 11.2$, 1.5 Hz), 7.20–7.52 (5 ArH, m); IR (CHCl₃) v 3170 (w), 2250 (C≡C, w), 1465 (s), 1382 (m), 910 (C=CH₂ bend, s), 725 (s), 650 (s), cm⁻¹; UV (95% EtOH) λ_{max} 327 nm (sh), 311; MS exact mass, m/e 276.1868 (calcd for C₂₁H₂₄, 276.1879); MS, m/e 276 (4, M), 261 (6), 203 (12), 178 (10), 155 (6), 154 (34), 153 (13), 152 (12), 144 (17), 130 (10), 129 (base), 128 (6), 122 (13), 115 (9), 109 (8), 105 (31), 102 (7), 95 (9), 93 (6), 91 (14), 81 (5), 79 (5), 77 (22), 76 (5), 75 (7), 74 (5).

1-[(1,1'-Z,2'-E)-3'-Methyl-5'-iodopent-2'-en-4'-yn-1'-ylidene]-2methylene-6,6-dimethylcyclohexane (20e). To a solution of trienyne 20a (55 mg, 0.27 mmol) in dry THF (0.8 mL) was slowly added n-butyllithium (0.20 mL, 1.6 M in hexanes, 0.32 mmol) at -78 °C under a nitrogen atmosphere. After further stirring for 15 min at -78 °C, powdered iodine (78 mg, 0.31 mmol) was introduced as a THF solution (1 mL). After the bright yellow mixture was stirred for 1 h at -78 °C, the mixture was worked up in the usual fashion (H₂O quench followed by warming to room temperature; lbpe extraction; back wash with sodium thiosulfate solution, brine, and water; Na₂SO₄ drying). Removal of the solvent followed by chromatography (silica gel, lbpe) afforded 79 mg (89%) of pure iodo compound 20e as an oil: ¹H NMR δ 1.06 (C₆ 2CH₃, s), 1.89 (C₃, CH₃, d, $J \sim 1.5$ Hz), 4.63 (C₂ CH_E, d, $J \sim 2.8$ Hz), 5.03 (C₂ CH_Z, d of t, $J \sim 2.8$ Hz, 1.4 Hz), 6.02 (H₁, d, $J \sim 10.8$ Hz), 6.76 (H₂, d with fine structure, $J \sim 10.8$ Hz); UV (95% EtOH) λ_{max} 287 nm (ϵ 22 500); MS exact mass, m/e 326.0511 (calcd for C₁₅H₁₉I, 326.0533); MS, m/e 327 (2, M + 1), 326 (4, M), 253 (base), 201 (37), 199 (23), 183 (21), 173 (27), 159 (42), 157 (28), 143 (28), 142 (38), 141 (32), 131 (22), 129 (27), 128 (69), 126 (81), 119 (22), 115 (32), 105 (32), 91 (34).

1-[(1,1'-Z,2'-E)-6'-Hydroxy-3'-methylhex-2'-en-4'-yn-1'-ylidene]-2methylene-6,6-dimethylcyclohexane (20g). To a solution of trienyne 20a (100 mg, 0.5 mmol) in ether (2 mL) was added *n*-butyllithium (0.4 mL, 1.6 M, 0.64 mmol at -10 °C), and the solution was stirred for 20 min under nitrogen. To the trienynyllithium suspension was added THF (1.5 mL) and then dry paraformaldehyde (30 mg, 1.0 mmol) whereupon a mildly exothermic reaction occurred. After the mixture was stirred for 1 h at room temperature, the flask was placed in an oil bath and heated

under reflux for 3 h and then stirred overnight at room temperature. The reaction mixture was poured into cold water (20 mL) and then the aqueous layer was reextracted with ether $(3 \times 10 \text{ mL})$ and dried over sodium sulfate. After removal of the ether, the residue was chromatographed (medium-pressure LC, silica gel; lbpe/ether, 20:80) to afford the desired alcohol **20g** (94 mg, 82%): 1 H NMR δ 1.05 (6 H, C₆ 2CH₃, s), 1.89 (3 H, $C_{3'}$ CH₃, d, $J \sim 1.5$ Hz), 4.31 (2 H, 2 H₆', br s, $w \sim 1.5$ Hz), 4.60 (1 H, C₂ CH_E, d, $J \sim 2.8$ Hz), 5.03 (1 H, C₂ CH_Z, dt, $J \sim$ 2.8, 1.5 Hz), 6.04 (1 H, $H_{1'}$, d, $J \sim 11.2$ Hz), 6.76 (1 H, $H_{5'}$, d with fine structure, $J \sim 11.2$ Hz); IR (CHCl₃) v 3615 (OH, w), 3165, 2940, 2870, 2210 (C=C), 1465, 1390, 1250, 1000, 910 cm⁻¹ (C=CH₂ bend, s); UV (95% EtOH) λ_{max} 283 nm (ϵ 23 600); ¹³C NMR δ 154.5, 145.6, 115.7 $(C_{1,2,3})$, 134.4, 115.6, 113.8 $(C_{2',1'})$, exo-methylene carbon), 90.1, 85.8 $(C_{5',4'})$, 51.8, 41.5, 38.5, 37.1, 27.5, 23.2, 17.3 (remaining sp³ seven carbons); MS exact mass; m/e 230.1681 (calcd for C₁₆H₂₂O, 230.1672); MS, m/e 230 (1, M), 163 (5), 159 (5), 149 (12), 147 (9), 145 (5), 137 (14), 136 (5), 135 (10), 133 (7), 131 (6), 125 (5), 123 (13), 121 (13), 119 (9), 111 (11), 109 (16), 107 (12), 105 (12), 98 (6), 97 (18), 96 (8), 95 (27), 93 (14), 91 (13), 85 (12), 84 (8), 83 (24), 82 (12), 81 (51), 79 (11), 77 (7), 71 (23), 70 (14), 69 (base), 68 (12), 67 (17), 65 (5), 57 (40), 56 (11), 55 (43), 53 (9).

(7Z)-4,4,13,13,13-Pentadeuterio-9-ethynyl- β -ionyl Benzoate (23). The pentadeuterio- β -ionone 29 was converted with lithium acetylide to the corresponding pentadeuterio-9-ethynyl- β -ionol according to the method previously described.⁸ The pentadeuterio-9-ethynyl- β -ionol was then converted to its 7Z isomer and then in turn to the *cis*-benzoate 23 according to the method of Reischl.9 Purification of 23 was achieved by high-pressure LC (Whatman Partisil M-9 10/50, reverse-phase ODS-2, 100% CH₃CN, 2 mL/min flow rate). The deuterium content of the benzoate 23 could not be readily assessed from its ¹H NMR spectrum at 200 MHz in that the signals due to the allylic methylene (2 H₄, δ 1.95) and allylic methyl (C₅ CH₃, $\delta \sim 1.8$) overlap with the C₉ methyl ($\delta 1.98$) and the remaining ring methylene groups (2 H₂ and 2 H₃, δ 1.4–1.9), respectively. However, in the ¹³C NMR spectrum of 23, the signals at δ 31.8 and 22.5 assigned to C₄ and C₁₃, respectively, in the undeuterated compound, have nearly completely degenerated into background noise as a result of multiple deuterium substitution at these carbons. Finally, although a complete deuterium content analysis $(d_0-d_5 \text{ species})$ of 23 was not achieved by mass spectrometry due to intensity problems at the time of measurement, a ratio of the d_5 to d_4 species could be accurately determined to be 63.79% d_5 to 36.21% d_4 . Of significance is that the deuterium content in terms of the d_5 to d_4 ratio of 25 prepared from this same sample of 23 was precisely identical. The exact masses of the molecular ions were as follows: m/e 327.2187 (calcd for C₂₂H₂₁D₅O₂, 327.2247); m/e 326.2187 (calcd for $C_{22}H_{22}D_4O_2$, 326.2185)

1-[(1,1'-Z,2'-E)-3'-Deuteriomethylpent-2'-en-4'yn-1'-ylidene]-2-dideuteriomethylene-3,3-dideuterio-6,6-dimethylcyclohexane (25). The cis-benzoate 23 was reacted with DBU as described in the synthesis of 20a. The reaction was worked up as before and the product subjected to high-pressure LC purification (Whatman Partisil M-9 10/50, 100%Skellysolve B, 3.0 mL/min flow rate). Mass spectral analysis of 25 analogous to that described for its precursor was carried out. The ratio of d_5 to d_4 species determined for 25 was 63.79% d_5 to 36.21% d_4 , precisely identical with that obtained for its precursor 23. The exact masses of the molecular ions were as follows: m/e 205.1875 (calcd for C₁₅H₁₅D₅, 205.1880); m/e 204.1817 (calcd for C₁₅H₁₆D₄, 204.1875). In another series of experiments, ¹H NMR was utilized to determine the isotopic composition by integration of the signal attributable to residual protons attached to the methyl carbon at $C_{3'}$ (δ 1.88). No signal was observed for residual protons attached to the allylic ring carbon C_3 (δ 2.20) or the exocyclic methylene carbon at C₂ (δ 4.62 and 5.02). Accordingly, it is assumed that the deuterium content at these positions is >95% for four deuterium atoms and, therefore, integral data are omitted below. As internal standards, the side chain olefinic $(H_{1'} \text{ and } H_{2'})$ and acetylenic $(H_{5'})$ protons were used. For comparison, the undeuterated hydrocarbon 20a was analyzed. The integration results were as follows:

	C ₃ ′ CH ₃ or CH₂D	H'.	H₂′	H⁵.
20a	2.96 ± 0.24	0.96 ± 0.08	1.09 ± 0.09	0.95 ± 0.08
25	1.98 ± 0.09	0.92 ± 0.04	1.15 ± 0.07	0.93 ± 0.05

The integration data are the averages of three separate scans using the cut and weigh method; the uncertainties are standard deviations. Finally, 13 C NMR data fully support the presence of a single deuterium atom on the C₃, methyl group. This and related data are presented in the supplementary material. Thus, the conversion of 23 to 25 occurs with complete retention of five deuterium atoms in support of the proposed pathway.

4,4,13,13,13.13.Pentadeuterio- β -ionone (29). The pentadeuterio- β -

cyclocitral 27 was prepared according to the procedure of Dawson¹² and converted to the corresponding nitrile 28 according to the procedure of Byers.¹³ To a solution of methyllithium (7.2 mmol) in dry THF (1 mL) was added slowly a solution of 28 (259 mg, 1.43 mmol) in dry THF (3.3 mL) at 0 °C. After the solution was stirred at 0 °C for 30 min, a mixture of sulfuric acid (1.2 mL, 3 M) and THF (3 mL) was added slowly. The reaction mixture was warmed to room temperature and stirred for a further 1.25 h. Water (25 mL) and methylene chloride (25 mL) were then added to the reaction mixture, and the organic layer was separated. The aqueous layer was reextracted with methylene chloride (25 mL). The organic extracts were combined, washed with water $(2 \times 50 \text{ mL})$ and saturated NaCl solution (50 mL), dried (MgSO₄), and concentrated under vacuum. The residue was chromatographed (silica gel; 15% ether/Skellysolve B) to give 29 (149 mg, 53%). The ¹H NMR spectra of 27 and 29 are indicative of >95% deuterium incorporation; these spectra are compared with the data for their unlabeled counterparts in the supplementary material.

(6Z,8E,12E)-10,11-Didehydro-18,14-retro-retinol (31b). A solution of trienyne 20a (100 mg, 0.50 mmol) in THF (0.8 mL) was slowly added to a solution of ethylmagnesium bromide (0.22 mL, 2.94 M, 0.64 mmol) in THF (1 mL). After the addition was complete, the mixture was heated at 55 °C for 1.5 h. The Grignard solution was added dropwise via cannula to a mixture of the (E)-bromide 30a (140 mg, 0.50 mmol) and tetrakis(triphenylphosphine)palladium (30 mg, 0.025 mmol) in benzene (2 mL). The mixture was stirred for 2 h at room temperature and then for 10 h at 60 °C. After cooling to room temperature, the mixture was quenched with cold water. The organic layer was separated, and the water layer was further extracted with ether $(3 \times 5 \text{ mL})$. The combined extracts were washed with brine and water. After the solution was dried (MgSO₄) and concentrated (vacuum), the resulting crude residue was passed through a short alumina column (8:2 lbpe/ether) and then concentrated in vacuo to afford 157 mg of the silvl ether 31a. To the crude silyl ether 31a was added freshly prepared tetra-n-butylammonium fluoride (2 mL, 1 M in THF, 2.0 mmol) at room temperature under nitrogen. The solution was stirred for 4 h and then poured into 10 mL of brine and extracted with ether. The separated aqueous layer was reextracted with ether (10 mL) and then the combined ether extracts were washed successively with 15 mL each of 1 M HCl, saturated aqueous NaHCO₃, and water. Upon drying (MgSO₄), filtering, and concentrating, the crude product was subjected to silica gel column chromatography (70:30 lbpe/ether). Combination and concentration of appropriate fractions afforded 75 mg (69%) of pure alcohol 31b. The corresponding Z isomer 32b was not detected as a byproduct: ¹H NMR δ 1.06 (C_{16,17} 2CH₃, s), 1.88 (C₂₀ CH₃, d, $J \sim 1.5$ Hz), 1.90 (C₁₉ CH₃, d, $J \sim 1.5$ Hz), 2.31 (2 H₁₄, t, $J \sim 6.5$ Hz), 3.66 (2 H₁₅, t, $J \sim 6.5$ Hz), 4.67 (H_{18E}, d, $J \sim 2.9$ Hz), 5.07 (H_{18Z}, dt, $J \sim 2.9$, 1.5 Hz), 5.47 (H₁₂, br q, $J \sim 1.5$ Hz), 6.10 (H₇, d, $J \sim 11.3$ Hz), 6.73 (H₈, dq, $J \sim 11.3$, 1.5 Hz); ¹³C NMR δ 153.7, 146.8, 145.6, 116.9 (C_{5,6,9,13}), 133.2, 115.9, 113.7, 107.8 (C_{7,8,12,18}), 97.3, 86.3 (C_{10,11}), 60.4 (C₁₅), 41.7, 41.5, 37.1 $(C_{2,3,4})$, 38.4 (C_1) , 27.6 $(C_{16,17})$, 23.2, 19.2, and 17.6 $(C_{14,19,20})$; IR (neat) ν_{max} 3360 (s, br, OH), 2180 (w, C=C), 900 (s, C=CH₂), other prominent bands at 2940, 1632, 1613, 1440, 1385, 1290, 1045 cm⁻¹; UV (95% EtOH), λ_{max} 311 nm (ϵ 27 600), 327 (sh, ϵ 22 500); MS exact mass, m/e284.2137 (calcd for $C_{20}H_{28}O$, 284.2141); MS, m/e 285 (18, M + 1), 284 (72, M), 269 (base), 253 (21), 239 (8), 223 (10), 215 (10), 209 (9), 197 (20), 183 (28), 171 (13), 169 (24), 159 (27), 143 (23), 137 (28), 131 (23), 119 (37), 105 (40), 91 (52), 77 (29), 69 (42), 55 (45).

(6Z,8E,12Z)-10,11-Didehydro-18,14-retro-retinol (32b). The trienyne 20a was coupled with the (Z)-bromide 30b in precisely the same manner as described for the E isomer 30a (see preceding section). The chromatographically pure alcohol 32b was obtained in 63% yield from a silica gel column with lbpe/ether (60:40). Further elution with ether gave the E isomer 31b in 6.2% yield: ¹H NMR δ 1.05 (C_{16,17} 2CH₃, s), 1.77 (C₂₀ CH₃, d, $J \sim 1.5$ Hz), 1.87 (C₁₉ CH₃, d, $J \sim 1.5$ Hz), 2.50 (2 H₁₄, t, $J \sim 6.3$ Hz), 3.67 (2 H₁₅, t, $J \sim 6.3$ Hz), 4.62 (H_{18E}, d, $J \sim 3.0$ Hz), 5.02 (H_{18Z}, dt, $J \sim 3.0$, 1.5 Hz), 5.47 (H₁₂, br s, $W \sim 4.5$ Hz), 6.03 (H₇, d, $J \sim 11.3$ Hz), 6.63 (H₈, dq, $J \sim 11.3$, 1.5 Hz); ¹³C NMR δ 153.7, 147.2, 145.6, 116.9 ($C_{5,6,9,13}$), 133.2, 115.9, 113.7, and 108.1 ($C_{7,8,12,18}$), 96.8, 86.3 ($C_{10,11}$), 60.9 (C_{15}), 41.5, 38.1, 37.2 ($C_{2,3,4}$), 38.5 (C_1), 27.6 ($C_{16,17}$), 23.2, 23.0, 17.6 ($C_{14,19,20}$); IR (neat) ν_{max} 3360 (s, br, OH), 2180 (w, C=C), 896 (s, C=CH₂), other prominent bands at 2930, 2870, 1630, 1440, 1385, 1040 cm⁻¹; UV (95% EtOH) λ_{max} 311 nm (ϵ 34 800), 326 (sh, ϵ 28 500); MS exact mass, m/e 284.2141 (calcd for $C_{20}H_{28}O$, 284.2141); MS, m/e 285 (10, M + 1), 284 (41, M), 269 (93), 253 (base), 239 (14), 223 (16), 213 (21), 209 (15), 197 (38), 183 (43), 173 (16), 169 (39), 159 (73), 155 (39), 143 (43), 133 (57), 119 (69), 105 (78), 91 (94), 77 (57), 69 (73), 55 (90).

1-[(1,1'-Z,2'-E,4'-E)-6'-Hydroxy-3'-methylhexa-2',4'-dien-1'-ylidene]-2-methylene-6,6-dimethylcyclohexane (33a). A solution of LiAlH₄ (40 mg, 1.1 mmol) in dry THF (2 mL) was cooled to 0 °C while propargyl alcohol 20g (50 mg, 0.2 mmol) in THF (1 mL) was added dropwise. The mixture was warmed to room temperature and then was heated under reflux for 3 h. After cautious addition of water, the reaction product was isolated by extraction with ether. The dried ether solution was concentrated and then the resulting residue was passed through a short silica gel column (50:50 lbpe/ether) to afford the tetraenol 33a in essentially quantitative yield: ¹H NMR δ 1.10 (6H, C₆ 2CH₃, s), 1.83 $(3 \text{ H}, \text{C}_{3'}\text{ CH}_3, \text{d}, J \sim 1.4 \text{ Hz}), 4.16 (2\text{H}, 2 \text{ H}_{6'}, \text{br d}, J \sim 6.2 \text{ Hz}), 4.64$ (1 H, C₂=CH_E, d, $J \sim 2.7$ Hz), 5.08 (1 H, C₂=CH_Z, dt, $J \sim 2.7$, 1.4 Hz), 5.81 (1 H, H_{5'}, dt, $J \sim 15.6$, 6.2 Hz), 6.23 (1 H, H_{1'}, d, $J \sim 10.8$ Hz), 6.35 (1 H, H_{4'}, d, $J \sim 15.6$ Hz), 6.57 (1 H, H_{2'}, d with fine structure, $J \sim 10.8$ Hz); IR (neat) $v_{\rm max}$ 3370 and 3320 (OH), 3380, 2940, 2870, 1635, 1620, 1460, 1100, 970, 900 (C=CH₂ bend , m) cm⁻¹; UV (95% EtOH) λ_{max} 287 nm (ϵ 27 200); MS exact mass; m/e 232.1836 (calcd for $C_{16}H_{24}O$, 232.1828); MS, m/e 233 (2, M + 1), 214 (2, M -H₂O), 217 (21), 201 (36), 199 (38), 187 (21), 159 (25), 157 (27), 147 (23), 145 (40), 143 (50), 141 (28), 133 (56), 131 (42), 129 (56), 128 (50), 119 (41), 117 (39), 116 (21), 115 (80), 109 (21), 107 (26), 105 (78), 95 (26), 93 (32), 91 (base), 81 (30), 79 (48), 77 (61), 69 (52), 67 (35), 57 (30), 55 (76).

1-[(1,1'-Z,2'-E,4'-E)-6-Formy1-3'-methylhexa-2',4'-dien-1'-ylidene]-2-methylene-6,6-dimethylcyclohexane (33b). Powdered MnO₂ (500 mg, 5.7 mmol) was added to a stirred solution of tetraenol 33a (50 mg, 0.2 mmol) in 3 mL of lbpe at 0 °C. After the mixture was stirred for 3 h under nitrogen at this temperature, the MnO₂ was removed by vacuum filtration through celite with lbpe. The lbpe was evaporated on the rotary evaporator and residue was subjected to high-pressure LC (partisil, 3% ethyl acetate/Skellysolve B, 4 mL/min flow rate) to give the pure aldehyde **33b** (40 mg, 81% yield): ¹H NMR δ 1.10 (6 H, C₆ 2CH₃, s), 1.93 $(3 \text{ H}, \text{C}_{3}' \text{ CH}_{3}, \text{d}, J \sim 1.9 \text{ Hz}), 4.72 (1 \text{ H}, \text{C}_{2} \text{ CH}_{E}, \text{d}, J \sim 2.6 \text{ Hz}), 5.17$ $(1 \text{ H}, \text{C}_2 \text{ CH}_2, \text{dt}, J \sim 2.6, 1.4 \text{ Hz}), 6.20 (1 \text{ H}, \text{H}_{5'}, \text{dd}, J \sim 7.5, 15.0$ Hz), 6.33 (1 H, H₁, d, $J \sim 11.3$ Hz), 6.97 (1 H, H₂, br d, $J \sim 11.3$ Hz), 7.20 (1 H, H₄', d, J ~ 15.0 Hz), 9.63 (CHO, d, J ~ 7.5 Hz); UV (95% EtOH) λ_{max} 342 nm (ε 30 000); IR (CCl₄) ν_{max} 2930 (s), 1685 (vs, CHO), 1595 (s, C=C), 1125, 970, 905 cm⁻¹ (C=CH₂ bend, m); MS exact mass, m/e 230.1674 (calcd for C₁₆H₂₂O, 230.1672); MS, m/e 231 (7, M + 1), 230 (42, M), 215 (58), 201 (40), 159 (85), 147 (25), 146 (25), 145 (base), 143 (33), 141 (27), 133 (80), 131 (80), 129 (43), 128 (41), 121 (26), 119 (48), 117 (49), 115 (54), 107 (35), 105 (77), 95 (40), 93 (36), 91 (base), 81 (34), 79 (48), 77 (69), 69 (42), 67 (28), 65 (33), 55 (60), 52 (41) 50 (35).

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Supplementary Material Available: ¹H NMR data for deuterium-labeled derivatives (3 pages). Ordering information is given on any current masthead page.